Calculating Percentage of PCNA- or TUNEL-positive Tumor or Ectopic Cells

After immunostaining tissue sections from three *SmoA1;Bmi1*^{+/+} and three *SmoA1;Bmi1*^{-/-} animals for PCNA or TUNEL as described in the Materials and Methods section, we photographed 10 random nonoverlapping high-power fields of tumor or ectopic tissue for each animal. If there were fewer than 10 high-power fields' worth of cells in a given *SmoA1;Bmi1*^{-/-} lesion, we photographed all ectopic cells present in the section. We then counted both the number of PCNA or TUNEL-positive nuclei per field and the total number of nuclei per field, and divided the number of positive nuclei by the total number of nuclei to obtain the percent positive nuclei. *P* values were calculated using the 2-tailed *t* test, assuming unequal variances.

Quantitative Real-time RT-PCR Primer Sets

Quantitative polymerase chain reaction was performed as described in the Materials and Methods section, using the following primer sets:

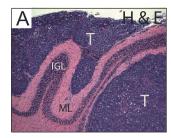
Actin: 5'-tgttaccaactgggacgaca-3' and 5'-tctcagctgtggtggtgaag-3'. Math1: 5'-tgcgctcactcacaaataag-3' and 5'-taacaacaacaatagtccgtgttc-3'. N-myc: 5'-gctgcggtcactagtgtgtc-3' and 5'-ggagaagcctcgctcttgat-3'. CyclinD1 [12]: 5'-ctctggctctgtgcctttct-3' and 5'-ccggagactcagagcaaact-3'.

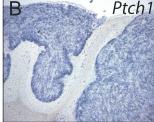
CyclinD2 [12]: 5'-ttcagcaggatgatgaagtga-3' and 5'-gagaaggggctag-cagatga-3'.

Supplementary References

[1] Oliver TG, Read TA, Kessler JD, Mehmeti A, Wells JF, Huynh TT, Lin SM, and Wechsler-Reya RJ (2005). Loss of patched and disruption of granule cell development in a pre-neoplastic stage of medulloblastoma. *Development* 132, 2425–2439.

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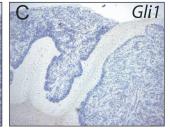


Figure W1. The Hh pathway is activated in SmoA1-induced medulloblastomas. (A) Hematoxylin and eosin staining of the cerebellum from a 7-week-old *SmoA1* mouse, demonstrating large tumor burden (T) outside the molecular layer (ML) and internal granular layer (IGL). (B–C) *In situ* hybridization for *Ptch1* (B) and *Gli1* (C) in nearby sections, demonstrating robust activation of the Hh pathway in tumors.

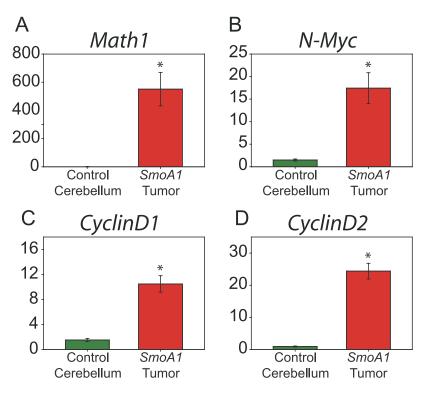


Figure W2. Medulloblastoma markers and Hh target genes are overexpressed in medulloblastomas. (A–D) Quantitative real-time RT-PCR reveals overexpression of *Math1* (A; 550-fold overexpression, P = .0097), N-Myc (B; 17-fold overexpression, P = .012), CyclinD1 (C; 11-fold overexpression, P = .0026), and CyclinD2 (D; 24-fold overexpression, P = .00064) in tumor-bearing cerebellum compared to control wild type cerebellum. N = 5 for each group; error bars indicate standard error of the mean; * indicates statistical significance.

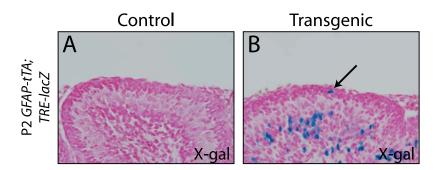


Figure W3. X-gal staining of 2-day-old *GFAP-tTA;TRE-lacZ* mouse cerebella. Overnight X-gal staining of P2 control (A) and *GFAP-tTA;TRE-lacZ* pups (B) revealed tTA activity in few cells scattered throughout the EGL at this stage. X-gal-stained cells in the EGL existed either singly or in small groups of cells.

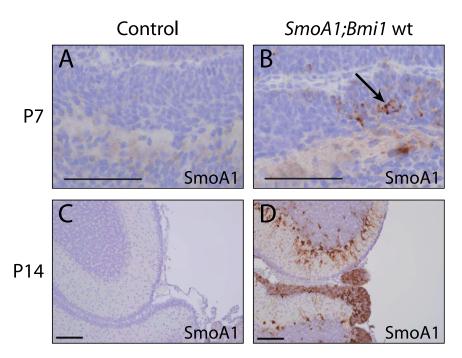


Figure W4. Immunostaining reveals focal SmoA1 expression in the mouse EGL at 7 and 14 days old. Detection of SmoA1 expression by immunostaining for the HA epitope tag revealed small clusters of SmoA1-positive cells in the EGL of 7-day-old pups (A, B, arrow), with only minor disruption of the normally smooth regular surface of the EGL. By 14 days old, whereas control EGL had begun to regress (C), readily identifiable SmoA1-expressing masses were already appreciable (D).

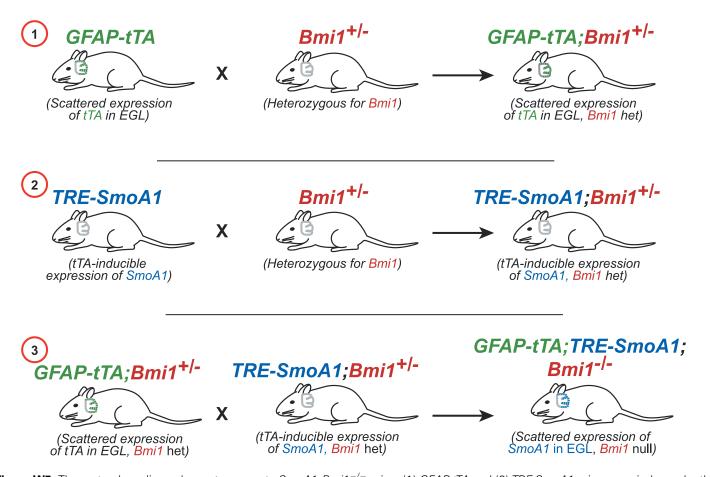


Figure W5. Three-step breeding scheme to generate SmoA1; $Bmi1^{-/-}$ mice. (1) GFAP-tTA and (2) TRE-SmoA1 mice were independently crossed with $Bmi1^{+/-}$ mice to generate GFAP-tTA; $Bmi1^{+/-}$ and TRE-SmoA1; $Bmi1^{+/-}$ mice, respectively. These progeny were then intercrossed (3) to generate GFAP-tTA; TRE-SmoA1; $Bmi1^{-/-}$ mice, designated SmoA1; $Bmi1^{-/-}$. A predicted 1 of 16 newborn pups from the final crosses (GFAP-tTA; $Bmi1^{+/-}$ × TRE-SmoA1; $Bmi1^{+/-}$) express SmoA1 on a Bmi1 null background (GFAP-tTA; TRE-SmoA1; $Bmi1^{-/-}$), but in reality, the actual proportion of mice obtained with this genotype is much lower because of the impaired viability of $Bmi1^{-/-}$ mice.

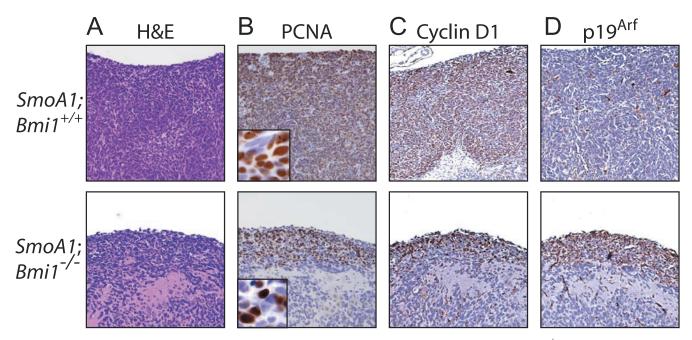


Figure W6. Cell cycle marker differences between intermediate tumor-like focus in P18 SmoA1; $Bmi1^{-/-}$ cerebellum and full-blown medulloblastoma in SmoA1; $Bmi1^{+/+}$ mice. (A) Hematoxylin and eosin staining of medulloblastoma SmoA1; $Bmi1^{-/-}$ and intermediate lesion (SmoA1; $Bmi1^{-/-}$). Histologic appearance of intermediate tumor-like lesion is similar to full-blown tumor, but the lesion is notably smaller. Shown in this image is the ectopic region with the maximal cross-sectional area. (B) PCNA stains both tumor and intermediate $Bmi1^{-/-}$ tumor-like focus, but stains fewer cells in the Bmi1-deficient lesion. Insets show higher magnification. Cyclin D1 expression is lower (C) and p19 Arf is elevated (D) in the SmoA1; $Bmi1^{-/-}$ lesion.

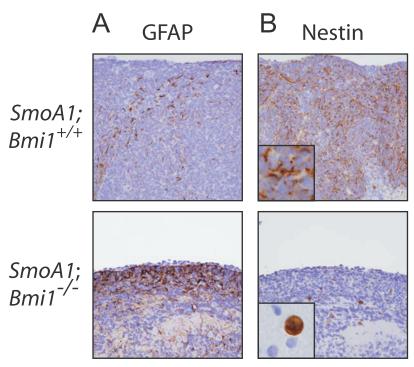


Figure W7. Increased glial density and loss of nestin expression in intermediate P18 SmoA1; $Bmi1^{-/-}$ tumor-like lesion. (A) Intense GFAP staining is seen in the outer region of the P18 SmoA1; $Bmi1^{-/-}$ tumor-like lesion, similar to the pattern seen in the ectopic regions of P21 animals. (B) Marked reduction of nestin-positive cells and alteration of subcellular nestin distribution is appreciable in SmoA1; $Bmi1^{-/-}$ mice at P18. Insets show higher magnification of tumor SmoA1; $Bmi1^{+/+}$ or molecular layer (SmoA1; $Bmi1^{-/-}$).